

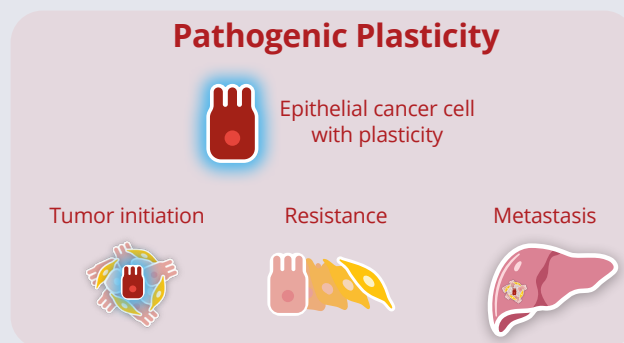
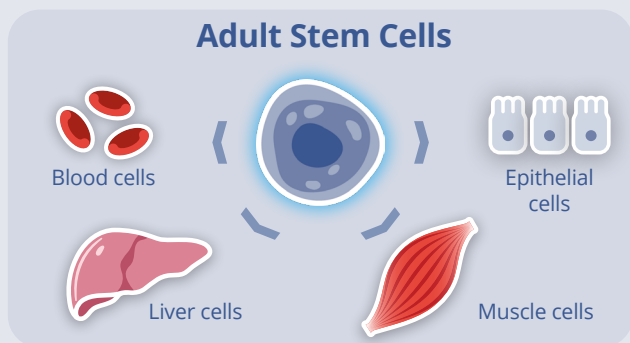
When Cancer Cells Acquire Pathogenic Plasticity, It Leads to Poor Patient Outcomes¹⁻⁴

Visit www.cancerswildcard.com to learn more.



Plasticity Is a Normal Part of Tissue Homeostasis, but It Can Be Exploited by Cancer^{1,2,5}

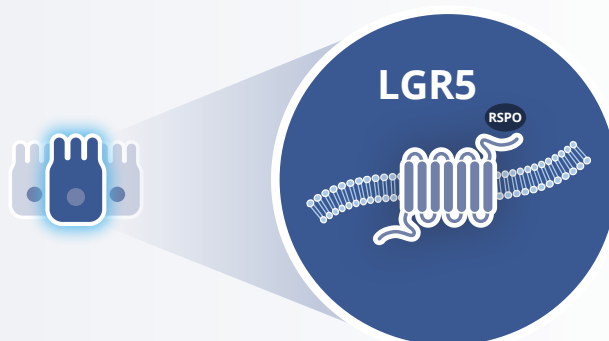
- **Plasticity:** characteristic of stem-like cells that enables self-renewal and differentiation into other cell phenotypes^{5,6}
 - Plasticity is typically reserved for a small subset of cells throughout the body called adult stem cells^{2,5,7,8}
 - Adult stem cells support normal homeostasis and injury repair (eg, cell replenishment)^{2,5,7,8}
- **Pathogenic plasticity:** cancer cells co-opting plasticity to initiate tumors, metastasize, and resist treatment⁵
 - Cancer cells can hijack regenerative capabilities to drive disease progression^{1,2,5}
 - Epithelial-mesenchymal transition (EMT) is an example of pathogenic plasticity that occurs in epithelial cancers^{1,5,9,10,a}



^aEMT is the process of epithelial cells losing their stationary characteristics (eg, cell-to-cell adhesion) and acquiring mesenchymal traits that facilitate motility and invasion.^{9,11}

In a Healthy Setting, Adult Stem Cells in Epithelial Tissue Express a Protein Called LGR5¹²

- LGR5 is a receptor for R-spondins (RSPO), growth factors involved in tissue homeostasis^{13,14}
 - RSPOs activate the Wnt/beta-catenin signaling pathway¹⁵
- **LGR5-positive cells support tissue regeneration¹²**
 - **In the intestine, LGR5-positive cells help replenish epithelial cells lost to shedding¹⁵⁻¹⁷**
- LGR5 is currently *not* an actionable biomarker^{12,18}
 - LGR5 protein detection is possible for preclinical purposes (eg, western blot, live cell imaging),¹⁹ but a suitable, well-validated antibody for human sample testing is not currently available^{12,18}



LGR5, Leucine-rich repeat-containing G protein-coupled receptor 5.



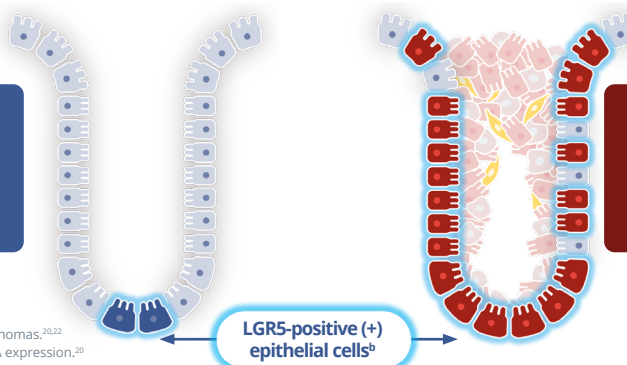
In a Pathogenic Setting, Cancer Cells Can Unlock LGR5 Expression, Which Enables Tumor Initiation^{12,20}

Colorectal tumorigenesis

- LGR5-positive cells are normally a small population of cells (eg, <10% of basal crypt cells in the intestinal epithelium)^{15,17,20}
- In the initial stages of colorectal tumorigenesis, the percentage of LGR5-positive cells has been shown to increase 10-fold (~70%),²⁰ which enables tumor cell proliferation and differentiation^{12,18,21}
- LGR5 is a Wnt-target gene and oncogenic mutations in the Wnt/beta-catenin pathway can lead to increased expression of LGR5¹⁸

Basal crypt of normal intestine

LGR5-positive cells are a small population of highly proliferative cells (<10%)^{15,17,20}



Tumor initiation^a

In colorectal tumorigenesis, the proportion of LGR5-positive cells can increase 10-fold²⁰

^aAdenomas and well/moderately differentiated adenocarcinomas.^{20,22}
^bbased on in situ hybridization (ISH) analysis of LGR5 mRNA expression.²⁰

Overexpression of LGR5 Has Been Associated With Poor Outcomes^{3,4,12,23-25}

- In colorectal cancer (CRC), LGR5 overexpression was associated with a ~30% reduction in disease-free survival (DFS; 1 year), a 3-fold increase in liver metastasis, and a 3-fold increase in disease recurrence (5 years)^{3,23,a}
- In head and neck cancer (H&N), LGR5 overexpression was associated with worse disease-specific survival (compared to low LGR5 expression) and markers of epithelial-mesenchymal transition (EMT)^{4,a}
- In lung cancer (NSCLC), LGR5 was overexpressed in tumor tissue compared with healthy adjacent tissue^{24,a}

Overexpression of LGR5 has been associated with^a

CRC^{3,23}



- Reduced DFS
- Liver metastasis
- Recurrence

H&N⁴



- Worse disease-specific survival (compared to low LGR5 expression)
- Markers of EMT^b

NSCLC²⁴



- Tumor tissue (compared with healthy adjacent tissue)

^aEvidence based on mRNA expression of LGR5.

^bMechanism of EGFR-treatment resistance.²⁶⁻²⁸

EGFR, epidermal growth factor receptor; EMT, epithelial-mesenchymal transition; mRNA, messenger RNA; NSCLC, non-small cell lung cancer.

See the full LGR5 story at CancersWildCard.com

1. Hanahan D. *Cancer Discov.* 2022;12(1):31-46; 2. Ge Y, Fuchs E. *Nat Rev Genet.* 2018;19(5):311-325; 3. Takahashi H, et al. *Ann Surg Oncol.* 2011;18(4):1166-1174;
4. Rot S, et al. *BMC Cancer.* 2019;19(1):155; 5. Torborg SR, et al. *Trends Cancer.* 2022;8(9):735-746; 6. Du X, et al. *Front Cell Dev Biol.* 2026;14:1752590; 7. National Institutes of Health. Stem cell basics. Accessed March 2026. <https://stemcells.nih.gov/info/basics/stc-basics>; 8. Brunet A, et al. *Nat Rev Mol Cell Biol.* 2023;24(1):45-62;
9. Makitie AA, et al. *Head Neck.* 2019;41(10):3712-3718; 10. Strauss R, et al. *Mol Ther.* 2012;20(5):887-897; 11. Thompson EW, et al. *Nat Rev Clin Oncol.* 2025;22(10):711-733;
12. Leung C, et al. *Trends Cell Biol.* 2018;28(5):380-391; 13. Carmon KS, et al. *Proc Natl Acad Sci U S A.* 2011;108(28):11452-11457; 14. Nagano K. *Jpn Dent Sci Rev.* 2019;55(1):80-87; 15. de Lau W, et al. *Genes Dev.* 2014;28(4):305-316; 16. Barker N, et al. *Cell Stem Cell.* 2010;6(1):25-36; 17. Barker N, et al. *Nature.* 2007;449(7165):1003-1007;
18. Mueller N, et al. *Immunother Adv.* 2025;5(1):Itaf017; 19. High PC, et al. *Cell Rep Med.* 2025;6(10):102363; 20. Martin ML, et al. *Cell Signal.* 2018;42:97-105;
21. Barker N, et al. *Nature.* 2009;457(7229):608-611; 22. Dow LE, et al. *Cell.* 2015;161(7):1539-1552; 23. Stanisavljevic L, et al. *Acta Oncol.* 2016;55(12):1425-1433;
24. Gao F, et al. *Transl Cancer Res.* 2019;8(1):203-211; 25. Xu L, et al. *Stem Cell Res Ther.* 2019;10(1):219; 26. Carosi F, et al. *Crit Rev Oncol Hematol.* 2026;221:105207; 27. Chhouri H, et al. *Cancers (Basel).* 2023;15(2):504; 28. Zhou J, et al. *J Exp Clin Cancer Res.* 2021;40(1):328.